ENGLEROMYCIN, A NEW CYTOCHALASAN FROM ENGLEROMYCES GOETZEI HENNINGS

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<u>Abstract</u>: The structure of engleromycin isolated from <u>Engleromyces</u> <u>goetzei</u> Hennings has been established as $\underline{1}$ by spectral techniques.

Engleromycin (<u>1</u>) is a new cytochalasan¹ which we have isolated from the fungus <u>Engleromyces goetzei</u> Hennings. It is a neutral substance, $C_{28}H_{35}NO_6$,² crystallizing in colourless needles from ethanol-H₂O, mp 226-228^o, $[\alpha]_D^{25}$ + 64^o(EtOH). On standing in pyridin and acetic anhydride <u>1</u> formed a diacetyl derivative.² Hydrogenation at atmospheric pressure (10 % Pd/C) gave a tetrahydro derivative.²

The structure of $\underline{1}$ was assigned mainly on the basis of spectral evidence. Fig. 1 shows the ¹H NMR spectrum and in Table 1 the chemical shifts are given. The spectrum is close to being identical to that published for cytochalasin D³ (<u>2</u>) (<u>cf</u>. Table 1) with a few exceptions caused by the following structural differences: <u>1</u> contains no acetyl group, only two double bonds, but three hydroxy groups, identified by the strong temperature dependence of their chemical shifts (10⁰



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corresponds to 50 Hz). The OH proton at 7.14 ppm must be strongly hydrogen bonded as seen from its low field chemical shift and the observed coupling constant. In addition to the mentioned differences the other distinct deviation from the spectrum of cytochalasin D (2) is the resonances at 3.996, 4.270 and 4.297 ppm, which are missing in the spectrum of 2. These shift values indicate that hydrogen is bonded through carbon to strongly electronegative atoms like oxygen. Decoupling experiments show that the proton at 4.270 ppm is coupled to the other two (tables 1 and 2) and that the proton at 4.297 ppm additionally is coupled to the OH proton at 7.14 ppm. The following moiety HO-CH-CH(O-)-CH(O-)- is thus indicated. The presence of three hydroxy groups and two carbonyl groups as observed in the ^{13}C NMR spectrum (Fig. 2 and Table 1) leaves, when taking into consideration that 1 contains six oxygen atoms, the following formulation for the moiety HO-CH-CH-CH-. A positive thiosulfate test confirmed the presence of an epoxide.⁴ Due to the

close spectral similarity of $\underline{1}$ and $\underline{2}$ and taking into consideration that $\underline{1}$ lacks an acetyl group and contains an epoxide at C(19)-C(20) in stead of the double bond of $\underline{2}$, makes it possible to formulate the structure of engleromycin as $\underline{1}$.

It is evident that it is the C(19)=C(20) double bond which is missing due to the observed coupling from the olefinic proton on C(13) to the proton on C(7). Since all cytochalasans isolated sofar¹ possess a C(19)=C(20) double bond it seems very reasonable to place the epoxy group at this position. But as an alternative it is not possible from the spectral data to exclude a structure with the epoxide group placed between C(20) and C(21), although this possibility from biogenetic reasons seems highly unlikely.

For epoxides the H-H <u>cis</u> coupling is 4.5 Hz, whereas the corresponding <u>trans</u> coupling is 3.1 Hz. In <u>1</u> the measured coupling is 2.5 Hz indicating a <u>trans</u> substitution. The coupling from the proton on C(20) to the proton on C(21) is very small, 0.8 Hz, indicating that the dihedral angle has to be close to 90° . This conformation is consistent with a conformation having a strongly hydrogen bonded C(21) OH group to the C(17) carbonyl group forming an eight-membered ring and, therefore, in agreement with placing the epoxy group at C(19) and C(20). The remaining coupling constants as given in Table 2 are very similar to those given for <u>2</u>.³

The ¹³C NMR spectrum (Fig. 2) shows all 28 resonances. The off-resonance decoupled spectrum gives multiplets as indicated in Table 1 corresponding to 31



protons attached directly to carbon. This together with the three OH groups and the NH group give the correct number of carbon and hydrogen $(C_{28}H_{35})$. All resonances except the ones at 60.90, 56.38 and 74.93 ppm relative to TMS can be assigned unambiguously from the chemical shifts given for <u>2</u> together with the multiplet structure in the off-resonance decoupled spectrum. Three lines were assigned by selective decoupling at the protons at C(19), C(20) and C(21).

In conclusion the structure of engleromycin $(\underline{1})$, with the uncertainty existting for the absolute configuration of the <u>trans</u>-substituted epoxide ring, can be formulated as $(7\underline{S}, 16\underline{S}, 18\underline{R}, 21\underline{S}) - 7, 18, 21$ -trihydroxy-19 $\underline{R}, 20\underline{S}$ -epoxy-16, 18-dimethy1 -10-pheny1-[11]cytochalasa-6(12), 13-diene-1, 17-dione <u>or</u> its 19 $\underline{S}, 20\underline{R}$ -epoxy isomer.

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References and Notes

- 1. M. Binder and C. Tamm, <u>Angew</u>. <u>Chem</u>. <u>85</u>, 369 (1973) and references cited therein.
- The compound was adequately characterized by spectral methods (IR, NMR and MS) and gave satisfactory high resolution mass spectral and/or combustion analytical data.
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- 5. The sample consisted of 15 mg of engleromycin in 0.4 ml pyridine-d₅ in a 5 mm sample tube. The sample was sealed and degassed by the usual freeze-pump-thaw technique. The spectra were recorded at 313 K and all chemical shifts are given at this temperature relative to TMS as internal standard. The spectra were measured on a Bruker HX-270 spectrometer with Fourier-transform system and a Nicolet 1180 computer.

The ¹H NMR spectra are an average of 10 scans, using 32K data points and a spectral range of 5000 Hz which corresponds to a digital resolution of 0.3 Hz. The ¹³C NMR spectra were obtained as an average of approximately 6000 scans using 32K data points and a spectral width of 17000 Hz giving a digital resolution of 1 Hz. The repetition time was 0.95 sec. and pulse width 4 µsec, corresponding to 60°. The decoupling power used in the selective decoupled spectra was adjusted to the minimum power necessary to observe the effect of the decoupling on C(11).

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